

# International Journal of Scientific Research in Dental and Medical Sciences



## www.ijsrdms.com

## Evaluate Enhanced Bone Regeneration by Human Adult Dental Pulp Stem Cells Combination with Scaffold: A Systematic Review and Meta-analysis

Sahar Raissi<sup>a</sup>, Mehrdad Barekatain<sup>b,\*</sup>, Nima Barekatain<sup>c</sup>, Shirin Zahra Farhad<sup>d</sup>, Mahdi Khadematolrasoul<sup>e</sup>, Sayeh Abbasnejad<sup>b</sup>

<sup>a</sup> Department of Prosthodontics, School of Dentistry, Kermanshah University of Medical Sciences, Kermanshah, Iran

<sup>b</sup> Department of Operative Dentistry, Faculty of Dentistry, Islamic Azad University, Isfahan (Khorasgan) Branch, Isfahan, Iran

c Faculty of Dentistry, Islamic Azad University, Isfahan (Khorasgan) Branch, Isfahan, Iran

ABSTRACT

<sup>d</sup> Department of Periodontics, Faculty of Dentistry, Islamic Azad University, Isfahan (Khorasgan) Branch, Isfahan, Iran

<sup>e</sup> Department of Periodontics, School of Dentistry, Centro Escolar University, Manila, Philippines

<sup>f</sup> Endodontics Department, School of Dentistry, Islamic Azad University, Tehran, Iran

## **ARTICLE INFO**

Article history:	Background and aim: The present study has tried to provide evidence in this field by summarizing the results of
Received 04 December 2023	animal studies because comprehensive results can lead to the decision of clinical trials in this field. Therefore, the
Received in revised form 01 February 2024	present study was conducted to evaluate enhanced bone regeneration by human adult dental pulp stem cells combined with scaffold.
Accepted 09 February 2024 Available online 15 February 2024	Material and methods: The present study was conducted based on PRISMA 2020-27-item checklist by giving the keywords stem cells, scaffold, bone regeneration and human dental pulp stem cells, all articles available in the international databases PubMed, Web of Science, Scopus, Science Direct, Web of Knowledge, EBSCO, Wiley, ISI,
Keywords: Bone Regeneration	Elsevier, Embase databases and Google Scholar search engine were reviewed until November 2023. STATA/MP. v17 software was used in this meta-analysis. Meta-analysis was performed using mean differences with 95% confidence interval in meta-analysis.
Dental Pulp Mesenchymal Stem Cells Scaffolds Stem Cells	<b>Results:</b> According to meta-analysis, the mean difference in bone regeneration between the experimental and control groups was 1.69 (MD:1.69; 95% CI $1.00 - 2.39$ , P<0.001). According to meta-analysis, a statistically significant difference was observed considering different groups of scaffolds on bone regeneration in combination with DPSC/SHED (MD:4.84; 95% CI 4.72–4.95, P<0.001).
Tissues	<b>Conclusions:</b> The present meta-analysis showed that dental pulp stem cells, along with scaffold, can increase new

bone formation and accelerate bone formation compared to the control group.

## 1. Introduction

One of the main and important challenges for orthopedic and craniofacial surgeons is the functional improvement of bone and complete reconstruction. Several complications have been reported for orthopedic and dental, the most important of which is the repair of traumatic and congenital defects and bone grafting.<sup>[11]</sup> Various methods have been reported for bone regeneration, including guided bone regeneration and bone grafting.<sup>[22]</sup> Since autogenous bone grafts are the best choice for bone regeneration, limited access to bone volume and donor site morbidity are reported complications.<sup>[31]</sup> Studies have used synthetic biomaterials and xenografts for bone graft scaffolds.<sup>[41]</sup> With the advancement of science and the development of tissue engineering, mesenchymal stem cells (MSCs) have been proposed, which can increase the

E-mail address: sayeh.abbasnejad@gmail.com

regeneration of bone tissue.<sup>[5, 6]</sup> MSCs have been considered suitable because of their self-renewal ability and multilineage differentiation.<sup>[7]</sup> MSCs can multiply at a high speed, have favorable paracrine, excellent bone-forming potential, and immunomodulatory properties like adult dental pulp stem cells (DPSCs) and stem cells from human exfoliated deciduous teeth (SHED).<sup>[8]</sup> About twenty years ago, DPSCs were introduced, and MSCs have shown good properties. DPSCs are among MSCs and can exogenously replace osteoblasts and multilineage differentiation. Some studies have shown that hDPSC and SHED induce bone regeneration equally.<sup>[9]</sup> The scaffold allows regeneration and facilitates growth factor binding. Based on the results of the studies, factors such as the type of stem cells and how they are combined with



<sup>\*</sup> Corresponding author. Sayeh Abbasnejad

Endodontics department, School of dentistry, Islamic azad university, Tehran, Iran https://doi.org/10.30485/IJSRDMS.2024.441841.1559

scaffold materials are effective in the success rate of bone regeneration. In this field, studies have been conducted that have been able to check the success rate of stem cells in vitro and in vivo using inorganic scaffold materials in bone regeneration.<sup>[10]</sup> So far, in vivo studies have been conducted to report evidence about the effect of DPSCs/SHED in bone regeneration. The present study has tried to provide evidence in this field by summarizing the results of animal studies because the comprehensive results can lead to the decision of clinical trials in this field. Therefore, the present study aimed to evaluate enhanced bone regeneration by human adult dental pulp stem cells combined with scaffold.

#### 2. Material and methods

#### Search strategy and Information sources

The present study was based on the PRISMA 2020-27-item checklist.<sup>[11]</sup> By giving the keywords stem cells, scaffold, bone regeneration, and human dental pulp stem cells, all articles available in the international databases PubMed, Web of Science, Scopus, Science Direct, Web of Knowledge, EBSCO, Wiley, ISI, Elsevier, Embase databases and Google Scholar search engine were reviewed until November 2023. In addition to this list of sources, the selected articles were screened to find relevant references. The search was done independently by two researchers to avoid bias.

The keywords studied based on the MeSH term were:

((((("Mesenchymal Stem Cells"[Mesh]) AND "Humans"[Mesh]) AND "Bone Regeneration"[Mesh]) OR "Bone Diseases"[Mesh]) OR "Oral and Maxillofacial Surgeons"[Mesh]) AND "Bone Transplantation"[Mesh].

#### Study selection criteria

## Inclusion criteria

Animal studies, used scaffold, availability of the full text of the article. Studies with incomplete results, case studies, case reports studies, and review articles were excluded.

#### Selection and data collection process

Two researchers independently collected data from the selected studies using a pre-prepared standard checklist to reduce reporting bias and errors. This checklist included Study specifications, clinical information, and study results.

#### Study risk of bias assessment

The CAMARADES checklist contained six independent items: randomization, controls, sample size calculation, published after peer review, outcome measure, and statement of potential conflict of interests. Studies that are more effective on the validity of the evaluation results regarding the therapeutic effect (moderate quality); Studies that affect the validity of the evaluation results about the therapeutic effect and change the evaluation results (low quality); Studies in which the results of the evaluation of the therapeutic effect are unclear (very low quality).

## Data analysis

STATA/MP. v17 software was used in this meta-analysis. Firstly, heterogeneity between studies was assessed by X2-based Q-tests and I2 tests (25%: low heterogeneity, 25-75%: moderate heterogeneity, and more than

75%: high heterogeneity) or was considered significantly heterogeneous (p<0.05). Meta-analysis was performed using mean differences with a 95% confidence interval in meta-analysis.

#### Study characteristics

Two hundred and thirty-eight animal samples were examined. In most of the selected studies, the treatment period was 8 weeks, and in three studies, it was 6 weeks. Other characteristics are reported in Table 1.

#### 3. Results

#### Study selection

In the initial search, 190 articles were found, and all articles were entered into EndNote.X8 software; in the first stage, by studying the titles of the articles, the number of 12 articles were deleted due to being repetitive. In the second step, by studying the abstract of 55 articles, 115 unrelated articles (based on the inclusion and exclusion criteria) were excluded from the study. In the third step, after carefully reading the full text of 55 articles, 42 articles were deleted due to inconsistency with the purpose of the study. Finally, 13 articles were used in this study (Fig. 1).

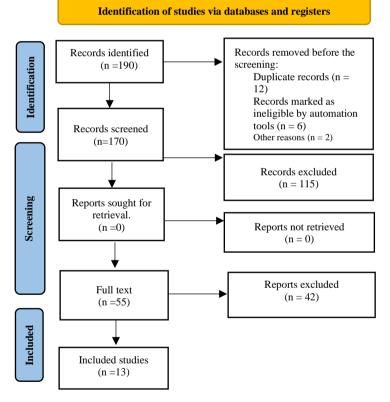


Fig. 1. PRISMA 2020 Checklist.

Table 1. Characteristics of the included studies.									
No.	Study. Years	Sample Size	Scaffolds Type	Experimental group	Defect Model of Bone	Follow-up (Weeks)			
1	Colorado et al., 2022 <sup>[12]</sup>	20	Polylactideco-glycolide	DPSC + Scaffold	Calvarial defect	10			
2	Vater et al., 2022 <sup>[13]</sup>	36	Mineralised collagen Matrix	DPSC + Scaffold	Critical mid- diaphyseal defect	6			
3	da Silva et al., 2022 <sup>[14]</sup>	50	Biphasic calcium phosphate	SHED + Scaffold	Calvarial defect	8			
4	Zhu et al., 2021 <sup>[15]</sup>	36	Bio-Oss—Collagen	DPSC + Scaffold	Calvarial defect	8			
5	Zhang et al., 2020 <sup>[16]</sup>	10	Tyrosine-derived polycarbonate	DPSC + Scaffold	Mandibular defect	23			
6	Salgado et al., 2020 <sup>[17]</sup>	4	Collagen–nanohydroxy apatite– phophoserine	DPSC + Scaffold	Subcutaneous implantation	8			
7	Bakopoulou et al., 2019 <sup>[18]</sup>	6	Biomimetic chitosan	DPSC + Scaffold	Subcutaneous implantation	10			
8	Huang et al., 2019 <sup>[19]</sup>	12	HNTs/GelMA hydrogels	DPSC + Scaffold	Calvarial defect	12			
9	Saha et al., 2019 <sup>[20]</sup>	20	Self-assembling $\beta$ -peptides	DPSC + Scaffold	Calvarial defect	6			
10	Jin et al., 2019 <sup>[21]</sup>	15	Puramatrix	DPSC + Scaffold	Mandibular bone defect	6			
11	Ansar et al., 2017 <sup>[22]</sup>	5	Alginate hydrogel with Cacl <sup>2</sup>	SHED + Scaffold	Subcutaneous implantation	8			
12	Wongsupa et al., 2017 <sup>[23]</sup>	18	PCL/BCP	SHED + Scaffold	Calvarial defect	15			
13	Fang et al., 2017 <sup>[24]</sup>	6	Collagen	SHED + Scaffold	Calvariae cranial defects	8			

According to meta-analysis, the mean difference in bone regeneration between the experimental and control groups was 1.69 (MD:1.69; 95% CI 1.00-2.39, P<0.001). According to these findings, bone regeneration in the experimental group was higher than in the control group. According to the test of group differences, a significant difference was observed between the groups under study (p<0.05). BV/TV had marginal effect (MD:5.01 mm<sup>3</sup>; 95% CI 2.55 mm 3-7.48 mm<sup>3</sup>, P<0.001), new bone formation (MD:2.08 mm<sup>2</sup>; 95% CI 0.70 mm2 - 3.47 mm2, P>0.001) and bone mineral density (MD:0.49 mg/cm3; 95% CI -0.60 mg/cm3-1.58 mg/cm3, P>0.001) shows no effect. bone formation showed highly significant effect (MD:2.20 mm<sup>2</sup>; 95% CI 0.82 mm2-3.58 mm2, P<0.001) (Fig. 2).

According to meta-analysis, a statistically significant difference was observed considering different groups of scaffolds on bone regeneration in combination with DPSC/SHED (MD:4.84; 95% CI 4.72- 4.95, P<0.001) (Fig. 3).

Study				Difference in Means with 95% Cl	Weigh (%)
BV/TV					
Zhu et al., 2021				3.60 [ -0.32, 7.52]	3.16
Jin et al., 2019		_		5.04 [ 1.12, 8.96]	3.16
Ansar et al., 2017				— 12.80 [ -0.92, 26.52]	0.26
Wongsupa et al., 2017				6.70 [ 0.82, 12.58]	1.41
Heterogeneity: $I^2 = 0.00\%$ , $H^2 = 1.00$	•			5.01 [ 2.55, 7.48]	
Test of $\theta_i = \theta_j$ : Q(3) = 2.05, p = 0.56					
Bone mineral density					
Vater et al., 2022	-			-0.43 [ -2.39, 1.53]	12.66
Zhang et al., 2020	-			1.10 [ -0.86, 3.06]	12.66
Huang et al., 2019				7.50 [ 3.58, 11.42]	3.16
Saha et al., 2019	-			-0.95 [ -2.91, 1.01]	12.66
Heterogeneity: $I^2 = 80.74\%$ , $H^2 = 5.19$	•			0.49 [ -0.60, 1.58]	
Test of $\theta_i = \theta_j$ : Q(3) = 15.58, p = 0.00					
Bone formation					
da Silva et al., 2022	-			3.08 [ 1.12, 5.04]	12.66
Salgado et al., 2020	-			1.11 [ -0.85, 3.07]	12.66
Bakopoulou et al., 2019				- 12.32 [ -1.40, 26.04]	0.26
Heterogeneity: $I^2 = 50.65\%$ , $H^2 = 2.03$	•			2.20 [ 0.82, 3.58]	
Test of $\theta_i = \theta_j$ : Q(2) = 4.05, p = 0.13					
New bone formation					
Colorado et al., 2022	-			1.37 [ -0.59, 3.33]	12.66
Fang et al., 2017	-			2.80 [ 0.84, 4.76]	12.66
Heterogeneity: $I^2 = 2.20\%$ , $H^2 = 1.02$	•			2.08 [ 0.70, 3.47]	
Test of $\theta_i = \theta_j$ : Q(1) = 1.02, p = 0.31					
Overall	•			1.69 [ 1.00, 2.39]	
Heterogeneity: $I^2 = 65.90\%$ , $H^2 = 2.93$					
Test of $\theta_i = \theta_j$ : Q(12) = 35.19, p = 0.00					
Test of group differences: $Q_b(3) = 12.48$ , p = 0.01					
	0	10	20	30	
ived offects inverse variance model					

Fixed-effects inverse-variance model

Fig. 2. The forest plot showed a sub-group meta-analysis of the overall effect of bone regeneration between the two groups.

Study						Difference in Means with 95% Cl		
Collagen + HA								(%)
Salgado et al., 2020		-				1.11 [ 0.52,	1.70]	3.79
Heterogeneity: $I^2 = 0.00\%$ , $H^2 = 1.00$		•				1.11 [ 0.52,	1.70]	
Test of $\theta_i = \theta_j$ : Q(0) = 0.00, p = .								
Collagen-containing scafold								
Vater et al., 2022						-0.43 [ -1.41,	0.55]	1.36
Zhu et al., 2021			-			3.59 [ 2.81,	4.37]	2.13
Jin et al., 2019			-			5.03 [ 4.64,	5.42]	8.53
Fang et al., 2017			-			2.81 [ 1.83,	3.79]	1.36
Heterogeneity: $I^2 = 97.31\%$ , $H^2 = 37.24$			•			4.02 [ 3.71,	4.33]	
Test of $\theta_i = \theta_j$ : Q(3) = 111.72, p = 0.00								
Hydroxyapatite -containing scafold								
Colorado et al., 2022		-				1.37 [ 0.78,	1.96]	3.79
da Silva et al., 2022						3.07 [ 2.87,	3.27]	34.12
Wongsupa et al., 2017			-			6.75 [ 6.36,	7.14]	8.53
Heterogeneity: $I^2 = 99.40\%$ , $H^2 = 165.70$			•			3.61 [ 3.44,	3.78]	
Test of $\theta_i = \theta_j$ : Q(2) = 331.40, p = 0.00								
Non-collagen- and non-HA-containing scafold								
Zhang et al., 2020						1.10 [ -0.47,	2.67]	0.53
Bakopoulou et al., 2019				-		12.30 [ 10.93,	13.67]	0.70
Huang et al., 2019						7.05 [ 6.85,	7.25]	34.12
Saha et al., 2019	_					-0.90 [ -2.86,	1.06]	0.34
Ansar et al., 2017						12.80 [ 11.43,	14.17]	0.70
Heterogeneity: $I^2 = 98.35\%$ , $H^2 = 60.50$			•	1		7.10 [ 6.91,	7.29]	
Test of $\theta_i = \theta_j$ : Q(4) = 241.99, p = 0.00								
Overall			1			4.84 [ 4.72,	4.95]	
Heterogeneity: I <sup>2</sup> = 99.26%, H <sup>2</sup> = 134.77								
Test of $\theta_i = \theta_j$ : Q(12) = 1617.21, p = 0.00								
Test of group differences: $Q_b(3) = 932.09$ , p = 0.00	·							
	-5	0	5	10	15	5		

Fixed-effects inverse-variance model

Fig. 3. The forest plot showed a sub-group meta-analysis of bone regeneration using different types of scaffolds between the two groups.

## 4. Discussion

Before animal studies are performed, all scaffolds used for bone regeneration have been analyzed in vitro.<sup>[25]</sup> Based on the present metaanalysis, DPSCs/SHED scaffolds were able to significantly increase bone regeneration. Also, a meta-analysis showed that dental pulp stem cells and scaffolds could significantly increase bone formation. As observed, there was a high heterogeneity between the studies, which indicates that the results of the present study should be interpreted with caution. The reason for this could be the difference in the cognitive methodology of the studies; some studies had a poor design, and the sample size was small. All these things can affect the results of the studies and the difference between the average results of the experimental and control groups. There is a need to conduct more studies with a higher sample size and appropriate and ethical cognitive methodology to confirm the current evidence and provide stronger evidence. A study has shown that SHED can increase mineralization capacity compared to DPSC.<sup>[26]</sup> Another study reported that the results of using SHED and DPSC in new bone formation were similar.<sup>[9]</sup> These findings are consistent with the present study's results; it was also observed that SHED and DPSC are similarly effective in bone regeneration. Also, the present study's findings indicate that the type of scaffold does not determine the effect of PSCs and SHED in bone

regeneration. Based on the available evidence, it is possible to extract dental pulp stem cells from unerupted wisdom teeth because these teeth are one of the most common methods of oral surgery.<sup>[27, 28]</sup>

Studies have also shown that using DPSC has been considered to improve the results of dental implants.<sup>[29]</sup> Currently, studies are investigating the effect of stem cells on bone regeneration, and strong evidence has not been provided.<sup>[30]</sup> Studies have shown that scaffold + dental stem cells are ineffective in new bone formation.[31-33] A systematic review and metaanalysis study reported that bone regeneration was significantly higher in the scaffold + hDPSC/SHED group than in the scaffold-only group.<sup>[34]</sup> Considering the differences in the findings of the studies, designing an ideal scaffold is challenging. However, the results of the present study show that integrating dental pulp stem cells with the ability of osteogenesis and the efficiency of the scaffold can increase the formation of new bone or, in other words, ossification. Studies that have been conducted in human clinical trials are very few. However, their findings show the positive effect of using dental pulp stem cells.<sup>[35-37]</sup> The present study had some limitations, such as the small sample size. Most of the studies did not observe the blinding of the experimental and control groups, which can affect the results of the studies, as well as the design of the studies regarding bone defect models, animal species, Gender, and recovery time.

## 5. Conclusion

The present meta-analysis showed that dental pulp stem cells and scaffolds can increase new bone formation and accelerate bone formation compared to the control group. Due to the ever-increasing elderly population and the economic burden, the primary need for bone tissue has increased. Based on the results of the present study and previous studies, dental pulp stem cells can be considered a promising option for ossification. More studies and clinical trials are needed to confirm these findings and the effectiveness of treatment based on dental pulp stem cells.

## **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### Acknowledgments

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

#### References

- Alqahtani AM. Guided tissue and bone regeneration membranes: A review of biomaterials and techniques for periodontal treatments. Polymers. 2023;15(16):3355. https://doi.org/10.3390/polym15163355.
- Hopper RA, Kapadia H, Susarla S, Bly R, Johnson K. Counterclockwise craniofacial distraction osteogenesis for tracheostomy-dependent children with Treacher Collins syndrome. Plastic and Reconstructive Surgery. 2018 Aug 1;142(2):447-57. https://doi.org/10.1097/PRS.000000000004606.
- [3] Sakkas A, Schramm A, Winter K, Wilde F. Risk factors for post-operative complications after procedures for autologous bone augmentation from different donor sites. Journal of Cranio-Maxillofacial Surgery. 2018;46(2):312-22. https://doi.org/10.1016/j.jcms.2017.11.016.
- [4] Haugen HJ, Lyngstadaas SP, Rossi F, Perale G. Bone grafts: which is the ideal biomaterial?. Journal of clinical periodontology. 2019:92-102. https://doi.org/10.1111/jcpe.13058.
- [5] Su P, Tian Y, Yang C, Ma X, Wang X, Pei J, et al. Mesenchymal stem cell migration during bone formation and bone diseases therapy.

International journal of molecular sciences. 2018;19(8):2343. https://doi.org/10.3390/ijms19082343.

- [6] Liu Y, Yang R, Shi S. Systemic infusion of mesenchymal stem cells improves cell-based bone regeneration via upregulation of regulatory T cells. Tissue Engineering Part A. 2015;21(3-4):498-509. https://doi.org/10.1089/ten.tea.2013.0673.
- [7] Visweswaran M, Pohl S, Arfuso F, Newsholme P, Dilley R, Pervaiz S, et al. Multi-lineage differentiation of mesenchymal stem cells–To Wnt, or not Wnt. The international journal of biochemistry & cell biology. 2015;68:139-47. https://doi.org/10.1016/j.biocel.2015.09.008.
- [8] Tatullo M, Marrelli M, Shakesheff KM, White LJ. Dental pulp stem cells: function, isolation and applications in regenerative medicine. Journal of tissue engineering and regenerative medicine. 2015;9(11):1205-16. https://doi.org/10.1002/term.1899.
- [9] Nakajima K, Kunimatsu R, Ando K, Ando T, Hayashi Y, Kihara T, et al. Comparison of the bone regeneration ability between stem cells from human exfoliated deciduous teeth, human dental pulp stem cells and human bone marrow mesenchymal stem cells. Biochemical and biophysical research communications. 2018;497(3):876-82. https://doi.org/10.1016/j.bbrc.2018.02.156.
- [10] Ercal P, Pekozer GG. A current overview of scaffold-based bone regeneration strategies with dental stem cells. Cell biology and translational medicine, volume 9: stem cell-based therapeutic approaches in disease. 2020:61-85. https://doi.org/10.1007/5584\_2020\_505.
- [11] Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. Bmj. 2021;372(71). https://doi.org/10.1136/bmj.n71.
- [12] Colorado C, Escobar LM, Lafaurie GI, Durán C, Perdomo-Lara SJ. Human recombinant cementum protein 1, dental pulp stem cells, and PLGA/hydroxyapatite scaffold as substitute biomaterial in critical size osseous defect repair in vivo. Archives of oral biology. 2022;137:105392. https://doi.org/10.1016/j.archoralbio.2022.105392.
- [13] Vater C, Männel C, Bolte J, Tian X, Goodman SB, Zwingenberger S. Effectiveness of dental pulp-derived stem cells and bone marrowderived mesenchymal stromal cells implanted into a murine critical bone defect. Current Stem Cell Research & Therapy. 2022;17(5):480-91. https://doi.org/10.2174/1574888X17666220215100732.
- [14] da Silva AA, Rinco UG, Jacob RG, Sakai VT, Mariano RC. The effectiveness of hydroxyapatite-beta tricalcium phosphate incorporated into stem cells from human exfoliated deciduous teeth for reconstruction of rat calvarial bone defects. Clinical oral investigations. 2022;26(1):595-608. https://doi.org/10.1007/s00784-021-04038-9.
- [15] Zhu Y, Wei SM, Yan KX, Gu YX, Lai HC, Qiao SC. Bovine-derived xenografts immobilized with cryopreserved stem cells from human adipose and dental pulp tissues promote bone regeneration: a radiographic and histological study. Frontiers in Bioengineering and Biotechnology. 2021;9:646690. https://doi.org/10.3389/fbioe.2021.646690.
- [16] Zhang W, Saxena S, Fakhrzadeh A, Rudolph S, Young S, Kohn J, et al. Use of human dental pulp and endothelial cell seeded tyrosine-derived polycarbonate scaffolds for robust in vivo alveolar jaw bone regeneration. Frontiers in Bioengineering and Biotechnology. 2020;8:796. https://doi.org/10.3389/fbioe.2020.00796.
- [17] Salgado CL, Barrias CC, Monteiro FJ. Clarifying the tooth-derived stem cells behavior in a 3D biomimetic scaffold for bone tissue engineering applications. Frontiers in bioengineering and biotechnology. 2020;8:724. https://doi.org/10.3389/fbioe.2020.00724.

- [18] Bakopoulou A, Georgopoulou A, Grivas I, Bekiari C, Prymak O, Loza K, et al. Dental pulp stem cells in chitosan/gelatin scaffolds for enhanced orofacial bone regeneration. Dental Materials. 2019;35(2):310-27. https://doi.org/10.1016/j.dental.2018.11.025.
- [19] Huang K, Ou Q, Xie Y, Chen X, Fang Y, Huang C, et al. Halloysite nanotube based scaffold for enhanced bone regeneration. Acs Biomaterials Science & Engineering. 2019;5(8):4037-47. https://doi.org/10.1021/acsbiomaterials.9b00277.
- [20] Saha S, Yang XB, Wijayathunga N, Harris S, Feichtinger GA, Davies RP, et al. A biomimetic self-assembling peptide promotes bone regeneration in vivo: A rat cranial defect study. Bone. 2019;127:602-11. https://doi.org/10.1016/j.bone.2019.06.020.
- [21] Jin Q, Yuan K, Lin W, Niu C, Ma R, Huang Z. Comparative characterization of mesenchymal stem cells from human dental pulp and adipose tissue for bone regeneration potential. Artificial cells, nanomedicine, and biotechnology. 2019;47(1):1577-84. https://doi.org/10.1080/21691401.2019.1594861.
- [22] Ansari S, Chen C, Hasani-Sadrabadi MM, Yu B, Zadeh HH, Wu BM, et al. Hydrogel elasticity and microarchitecture regulate dental-derived mesenchymal stem cell-host immune system cross-talk. Acta biomaterialia. 2017;60:181-9. https://doi.org/10.1016/j.actbio.2017.07.017
  - https://doi.org/10.1016/j.actbio.2017.07.017.
- [23] Wongsupa N, Nuntanaranont T, Kamolmattayakul S, Thuaksuban N. Assessment of bone regeneration of a tissue-engineered bone complex using human dental pulp stem cells/poly (ε-caprolactone)-biphasic calcium phosphate scaffold constructs in rabbit calvarial defects. Journal of Materials Science: Materials in Medicine. 2017;28:1-4. https://doi.org/10.1007/s10856-017-5883-x.
- [24] Fang TJ, Wang DH, Wang CY, Poongodi R, Liou NH, Liu JC, et al. Osteogenic prospective of deriving human dental stem cells in collagen matrix boost. Journal of Materials Science: Materials in Medicine. 2017;28:1-0. https://doi.org/10.1007/s10856-017-6001-9.
- [25] El-Rashidy AA, Roether JA, Harhaus L, Kneser U, Boccaccini AR. Regenerating bone with bioactive glass scaffolds: A review of in vivo studies in bone defect models. Acta biomaterialia. 2017;62:1-28. https://doi.org/10.1016/j.actbio.2017.08.030.
- [26] Wang X, Sha XJ, Li GH, Yang FS, Ji K, Wen LY, et al. Comparative characterization of stem cells from human exfoliated deciduous teeth and dental pulp stem cells. Archives of oral biology. 2012;57(9):1231-40. https://doi.org/10.1016/j.archoralbio.2012.02.014.
- [27] Dodson TB, Susarla SM. Impacted wisdom teeth. BMJ clinical evidence. 2010;2010:1302.
- [28] Kaminishi RM, Lam PS, Kaminishi KS, Marshall MW, Hochwald DA. A 10-year comparative study of the incidence of third molar removal in the aging population. Journal of oral and maxillofacial surgery. 2006;64(2):173-4. https://doi.org/10.1016/j.joms.2005.10.007.
- [29] Ito K, Yamada Y, Nakamura S, Ueda M. Osteogenic potential of effective bone engineering using dental pulp stem cells, bone marrow

stem cells, and periosteal cells for osseointegration of dental implants. International Journal of Oral & Maxillofacial Implants. 2011;26(5).

- [30] Polo-Corrales L, Latorre-Esteves M, Ramirez-Vick JE. Scaffold design for bone regeneration. Journal of nanoscience and nanotechnology. 2014;14(1):15-56. https://doi.org/10.1166/jnn.2014.9127.
- [31] Annibali S, Cicconetti A, Cristalli MP, Giordano G, Trisi P, Pilloni A, et al. A comparative morphometric analysis of biodegradable scaffolds as carriers for dental pulp and periosteal stem cells in a model of bone regeneration. Journal of Craniofacial Surgery. 2013;24(3):866-71. https://doi.org/10.1097/SCS.0b013e31827ca530.
- [32] Goncalves F, de Moraes MS, Ferreira LB, Carreira AC, Kossugue PM, Boaro LC, et al. Combination of bioactive polymeric membranes and stem cells for periodontal regeneration: in vitro and in vivo analyses. PLoS One. 2016;11(3):e0152412. https://doi.org/10.1371/journal.pone.0152412.
- [33] Jahanbin A, Rashed R, Alamdari DH, Koohestanian N, Ezzati A, Kazemian M, et al. Success of maxillary alveolar defect repair in rats using osteoblast-differentiated human deciduous dental pulp stem cells. Journal of Oral and Maxillofacial Surgery. 2016;74(4):829-e1. https://doi.org/10.1016/i.joms.2015.11.033.
- [34] Namjoynik A, Islam MA, Islam M. Evaluating the efficacy of human dental pulp stem cells and scaffold combination for bone regeneration in animal models: A systematic review and meta-analysis. Stem cell research & therapy. 2023;14(1):132. https://doi.org/10.1186/s13287-023-03357w.
- [35] d'Aquino R, De Rosa A, Lanza V, Tirino V, Laino L, Graziano A, et al. Human mandible bone defect repair by the grafting of dental pulp stem/progenitor cells and collagen sponge biocomplexes. Eur Cell Mater. 2009;18(7):75-83.
- [36] Hernández-Monjaraz B, Santiago-Osorio E, Ledesma-Martínez E, Alcauter-Zavala A, Mendoza-Núñez VM. Retrieval of a periodontally compromised tooth by allogeneic grafting of mesenchymal stem cells from dental pulp: a case report. Journal of International Medical Research. 2018;46(7):2983-93. https://doi.org/10.1177/0300060518773244.
- [37] Tanikawa D, Pinheiro CC, Almeida MC, Oliveira CR, Coudry RD, Rocha DL, et al. Deciduous dental pulp stem cells for maxillary alveolar reconstruction in cleft lip and palate patients. Stem Cells International. 2020;2020. https://doi.org/10.1155/2020/6234167.

How to Cite this Article: Raissi S, Barekatain M, Barekatain N, Farhad SZ, Khadematolrasoul M, Abbasnejad S. Evaluate Enhanced Bone Regeneration by Human Adult Dental Pulp Stem Cells Combination with Scaffold: A Systematic Review and Meta-analysis. International Journal of Scientific Research in Dental and Medical Sciences. 2024;6(1):17-23.

https://doi.org/10.30485/IJSRDMS.2024.441841.1559.